Short telomeres in patients with chronic schizophrenia who show a poor response to treatment

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Objective: Telomere shortening has been observed in many human diseases, including atherosclerosis, cancer, aging syndromes, Alzheimer disease and vascular dementia. The present study aimed to investigate the mean telomere lengths of patients with schizophrenia.

Methods: We analyzed the lengths of telomeric DNA, comparing 2 groups of patients with schizophrenia (34 good responders and 34 poor responders). A control group of 76 healthy volunteers was also included. Blood samples were obtained, and telomere length was measured by Southern blot analysis on the mean length of terminal restriction fragment (TRF).

Results: Compared with the control group, a significant amount of telomere shortening was found in peripheral blood leukocytes from patients with schizophrenia who experienced poor response to antipsychotics ($p < 0.001$). Conclusion: Shortened telomere length in chronic schizophrenia may be a trait marker caused by oxidative stress, and the ensuing cellular dysfunction may be a factor contributing to the progressive deterioration in treatment-resistant schizophrenia.

Introduction

Telomeres, the special structures at the ends of human chromosomes, are composed of repetitive DNA and DNA-binding proteins. They serve as caps with several functions, including protecting the ends of chromosomes, preventing chromosome fusion, facilitating chromosome segregation, distinguishing a chromosome end from a double strand break in the genomic DNA and maintaining genome stability.$^1$ Because of the limiting nature of linear DNA replication mechanisms, the telomeric DNA shortens in each round during cell division.$^2$ The telomeres in fibroblasts of children are longer than those in older adults (~9 kilobases [kb] v. ~4–6 kb).$^3$ However, in the human germ line, telomeres are 10–20 kb,$^4$ and it is thought that the lengths are maintained by the balanced interaction between the enzyme telomerase and various telomere-associated proteins.$^5$ In contrast to the germ line and early embryonic cells, most somatic cells do not express telomerase.$^6$ It is thought that normal cells respond to critically shortened telomeres, which are presumably
dysfunctional, by undergoing cellular senescence. Human cells senesce when the telomeres reach an average length of 4–7 kb. Telomere shortening has been observed in many studies including tissues with high cell turnover and in the pathogenesis of several diseases. Aging-related telomere shortening has been detected in cell and tissue types such as fibroblasts, leukocytes, vascular tissues and liver and kidney tissue. Shortened telomeres have been reported in studies of human diseases that include atherosclerosis, cancer, aging syndromes, vascular dementia and Alzheimer disease.

In the present study, we investigated the relation between mean telomere length and treatment response of inpatients with chronic schizophrenia.

Methods

We recruited 68 inpatients with schizophrenia from Kai-Suan Psychiatric Hospital, Kaohsiung (a major psychiatric centre in Taiwan). The study was approved by the Institutional Review Board at the Kai-Suan Psychiatric Hospital and conducted in accordance with the Declaration of Helsinki. The Structured Clinical Interview for the Diagnostic and Statistical Manual of Mental Disorders, fourth edition (DSM-IV) was used for the diagnosis. All patients met the criteria for a diagnosis of schizophrenia at admission, determined by 2 experienced psychiatrists. Subjects were physically healthy, and all laboratory parameters were within normal limits. They had no DSM-IV criteria of alcohol or substance abuse or neurodegenerative disorder on axis I. They did not present signs of disorientation to time, place or person, nor did they show any sign of impairment in immediate, recent or long-term memory. These patients were capable of performing routine daily activities, participating in occupational and recreational therapy and independently managing their personal affairs, such as laundry or money spending. We obtained their informed consent after the procedures had been fully explained. The average age of patients was 38 (range 19–59) years. We evaluated their clinical state at baseline and recorded findings based on the Brief Psychiatric Rating Scale (BPRS) and Global Assessment of Functioning scores (GAF). The patients were classified into 2 groups by a median split of GAF scores. The good responders comprised 34 patients who scored higher than 40 on the GAF (mean 48.4, standard deviation [SD] 6.6), and the poor responders comprised 34 patients who scored less than or equal to 40 (mean 31.2, SD 6.2). Age-matched healthy volunteers were recruited as control subjects. Control subjects had no psychiatric history, were not taking any psychotropic medication and did not have any first-degree relative with schizophrenia.

Telomere length measurement (terminal restriction fragment [TRF] assay) was performed according to the standard protocol, as previously described. Leukocyte DNA was extracted from peripheral blood samples. Five micrograms of DNA were digested overnight with Hinf I restriction endonuclease (Takara Bio Inc., Otsu, Shiga, Japan) to produce a terminal restriction fragment, an approximate simulation of the telomeric DNA. Equal quantities of digested DNA were loaded on a 0.6% agarose gel at 33 V for 4–5 hours in 1 x tris-acetate-EDTA buffer (40 mM Tris-acetate, 1 mM ethylenediaminetetraacetic acid). After electrophoresis, the DNA was denatured directly on the gel by treatment with denaturing and neutralizing buffers and then transferred to nylon membranes (Hybond-N+; Amersham, Little Chalfont, UK) for Southern blotting. The membranes containing transferred DNA were hybridized with a biotin-labelled oligonucleotide probe specific for the telomeric DNA repeat sequence (TTAGGG). Hybridization was performed at 37°C for 16–18 hours in a hybridization buffer (DIG Easy Hyb; Roche Applied Science, Mannheim, Germany). Unbound probe was removed by 2 washes in 0.15 M sodium chloride/0.015 M sodium citrate at 37°C and 2 washes with 15 mM sodium chloride/1.5 mM sodium citrate. Telomeric smears were detected with a Biotin Luminescent Detection Kit (Roche Applied Science). The mean TRF lengths were analyzed with the Photo-Capt, Version 99.03 software (Photo-Print IP-008-SD; Vilber Lourmat, Marne-la Vallée, France) by integrating the signal intensity of the TRF smear on the film as a function of its mean molecular weight, which was determined based on the standard biotin-labelled Hind III–digested markers of known molecular weight.

The differences in mean TRF between good responders, poor responders and control subjects, with consideration of possible confounders, were assessed by analysis of covariance followed by Tukey’s multiple comparison procedure. All statistical analyses were performed with SAS statistical software for Windows (SAS 9.1, SAS Institute Inc., NC).

Results

The demographic and clinical characteristics of the sample are presented in Table 1. There were no statistically significant differences in age (p = 0.86) or sex (p = 0.29) among these 3 groups, nor were there significant differences between schizophrenia subgroups in age of onset (p = 0.35), illness duration (p = 0.88) and medication dosage in chlorpromazine equivalent.

Table 1: Demographic and clinical characteristics of patients and control subjects

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Good responders</th>
<th>Poor responders</th>
<th>Control subjects</th>
<th>P value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, y</td>
<td>38.6 (7.5) [26–60]</td>
<td>37.3 (9.1) [19–51]</td>
<td>38.2 (10.9) [19–60]</td>
<td>0.86</td>
</tr>
<tr>
<td>Age of onset, y</td>
<td>23.1 (7.4) [15–49]</td>
<td>21.6 (6.6) [13–47]</td>
<td>NA</td>
<td>0.35</td>
</tr>
<tr>
<td>Illness duration, y</td>
<td>15.8 (7.8) [1–29]</td>
<td>16.0 (6.8) [1–29]</td>
<td>NA</td>
<td>0.88</td>
</tr>
<tr>
<td>CPZE, mg</td>
<td>580 (340) [199–931]</td>
<td>697 (431) [199–729]</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>BPRS total score</td>
<td>38.1 (8.0) [26–50]</td>
<td>44.1 (12.3) [21–71]</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>GAF score</td>
<td>48.4 (6.6) [31–62]</td>
<td>31.2 (6.2) [16–51]</td>
<td>NA</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>

BPRS = Brief Psychiatric Rating Scale; CPZE = chlorpromazine equivalent; F = female; GAF = Global Assessment of Functioning; M = male; NA = not applicable; SD = standard deviation. *Categorical data were analyzed with χ2 test, continuous data with 1-way analysis of variance or t-test.
lents ($p = 0.22$). Figure 1 shows TRFs of a representative Southern blot for samples obtained from schizophrenia patients. Average values of mean telomere lengths for the 3 groups were significantly different after controlling for age and sex (Table 2). Individuals in the poor responders group had the shortest mean TRF length (7.41, SD 0.97, range 5.95–9.69 kb), compared with the good responders group (8.88, SD 0.90, range 6.56–10.24 kb) and the control subjects (8.91, SD 1.36, range 5.97–11.14 kb) (Table 2). The mean telomere length of the poor responders was significantly shorter than that of the control subjects (Tukey’s multiple comparison procedure, $p < 0.001$, Table 2). In addition, comparison between good responders and poor responders also showed significant difference (Tukey’s multiple comparison procedure, $p < 0.001$, Table 2). There was no difference between the good responders and the control subjects (Tukey’s multiple comparison procedure, $p > 0.05$, Table 2). Interestingly, the TRF length was found to be inversely associated with age only in the control subjects; we found no association in the patient groups (Fig. 2). Mean telomere length in the control samples showed a net decrease with age of 79 base pairs (bp) yearly, assuming a constant rate of TRF loss with aging ($t = 11.95–0.079$, $p < 0.001$, Table 2). In addition, a net reduction with age of 79 bp yearly was found in the control group in this study. The observation is in accordance with published data that documented a loss of 31–84 bp yearly.21,22

Several studies, including ours, suggest that there is an inverse association between telomere length and age in healthy people. However, reduction of telomere lengths in patients with schizophrenia is not inversely associated with age. As well, no inverse association between telomere length and age was found in patients with probable Alzheimer dementia or in patients with stroke and related risk factors.23

### Discussion

Our results indicate that telomere length is shortened significantly in a subgroup of chronic schizophrenia patients who respond poorly to treatment. This is the first study, to our knowledge, to examine telomere lengths from peripheral blood leukocytes of patients with chronic schizophrenia.

We observed that the telomere lengths of the poor responders were the shortest among the 3 groups studied. On average, the mean telomere lengths of the poor responders were 1.5 kb shorter than those of age-matched control subjects. Additionally, a net reduction with age of 79 bp yearly was found in the control group in this study. The observation is in accordance with published data that documented a loss of 31–84 bp yearly.21,22

Fig. 2: Association between mean telomere restriction fragment and age in control subjects (open circles), good responders with schizophrenia (black circles) and poor responders with schizophrenia (triangles). The linear regression lines are given for the control subjects, good responders and poor responders, respectively. The slopes of the 3 groups are different by the test of homogeneity of slopes from the analysis of covariance ($p < 0.001$). Good responders slope $\beta = 0.020$, $F_{1,32} = 0.920$, $p = 0.35$. Poor responders slope $\beta = 0.007$, $F_{1,32} = 0.13$, $p = 0.72$. Control subjects slope $\beta = -0.080$, $F_{1,32} = 52.0$, $p < 0.001$.

### Table 2: Terminal restriction fragments of patients with schizophrenia and control subjects

<table>
<thead>
<tr>
<th>Measurement</th>
<th>Good responders* ($n = 34$)</th>
<th>Poor responders* ($n = 34$)</th>
<th>Control subjects* ($n = 76$)</th>
<th>$&lt;$ 40 y ($n = 27$)</th>
<th>$\geq$ 40 y ($n = 51$)</th>
<th>$&lt;$ 40 y ($n = 51$)</th>
<th>$\geq$ 40 y ($n = 57$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>TRF</td>
<td>8.88 (0.90)*</td>
<td>7.41 (0.97)*</td>
<td>8.91 (1.36)*</td>
<td>8.97 (1.27)</td>
<td>8.05 (1.14)</td>
<td>8.91 (1.19)</td>
<td>8.15 (1.37)</td>
</tr>
</tbody>
</table>

* $p < 0.001$. By analysis of covariance, controlled for age ($p = 0.18$, sex ($p = 0.53$), group x sex ($p = 0.95$), group x age ($p = 0.012$), group x sex x age ($p = 0.61$). Means with different letters are significantly different (according to Tukey’s multiple comparison procedure).
It was hypothesized that telomere erosion might serve as a biological clock that could count mitotic time and account for cell replicative senescence in culture. Loss of replicative capacity leads to cell growth arrest, which occurs not only after accumulated doubling populations with telomere shortening in culture but also as a consequence of subcytotoxic stress, such as mild chronic oxidative stress. Telomere shortening intensified by oxidative stress has been seen in induced conditions of cell cultures, such as chronic hyperoxia, as well as in fibroblasts from patients with Fanconi anemia and in peripheral blood leukocytes from patients with respiratory chain disorders, both of which diseases result in increased oxidative stress.

Several studies have demonstrated that reduction in antioxidant capacity and rise in oxygen free radicals might contribute to oxidative stress in schizophrenia. Recently, it has been shown that oxidative stress might cause mitochondrial dysfunction and altered brain metabolism in schizophrenia, which raises a possibility that increased oxidative stress, as shown by our observation of short telomeres, might exist in poor responders with chronic schizophrenia. This was supported by a report showing that oxidative DNA damage was 10 times greater in postmortem hippocampal of elderly patients with “poor-outcome” schizophrenia. Further studies are needed to establish the status and role of oxidative stress in poor responders with chronic schizophrenia and in the pathogenesis of schizophrenia.

In conclusion, we found that the treatment-resistant patients with chronic schizophrenia have significantly shortened telomeres, which may have been caused by oxidative stress that led to ensuing cellular dysfunction. The findings also point to shortened telomere lengths as a trait marker for chronic schizophrenia patients with poor response to treatment.

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Contributors: Drs. Yu and Cho contributed equally to the work. Drs. Yu acquired the data, which Drs. Chang, Lin and Cho analyzed. Drs. Yu, Lin and Cho wrote the article, which Drs. Chang and Cho reviewed. All authors gave final approval for publication.

References